

# THE ABSOLUTE CONFIGURATION OF THE NEW AMINO ACID 2-AMINO-4-METHYL-HEX-5-ENOIC ACID FROM A NEW GUINEA *BOLETUS*\*

EMERY GELLERT, BERTHOLD HALPERN and RICHARD RUDZATS

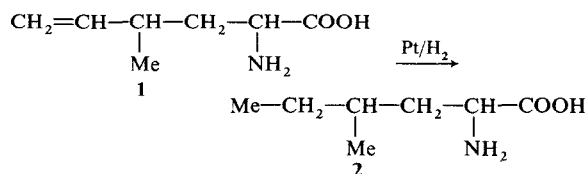
Department of Chemistry, University of Wollongong, Wollongong, N.S.W. 2500, Australia

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**Key Word Index**—*Boletus*; Boletaceae; amino acids; absolute configuration.

**Abstract**—The absolute configuration of the 2-amino-4-methyl-hex-5-enoic acid isolated from *Boletus* was shown to be 2*S*, 4*S*, by an unambiguous synthesis of its dihydro derivative from 2*S*-(−)-2-methylbutan-1-ol.

We have previously reported the isolation of 2-amino-4-methyl-hex-5-enoic acid (**1**) [1, 2] and shown that the asymmetric center at carbon-2 was *S* but could not establish the configuration at carbon-4 by physicochemical methods. We have now synthesized (2*S*,4*S*)-2-amino-4-methyl hexanoic acid (**2**) unambiguously from 2*S*-(−)-methylbutan-1-ol and acetylaminomalonate [3, 4], followed by resolution of the carbon-2 center of the acetylaminomethylhexanoic acid by hog kidney acylase I [5, 6]. The synthetic product was identical in all respects with the dihydro compound (**2**) obtained from the natural product (**1**) by catalytic hydrogenation. As catalytic hydrogenation of the Δ5 double bond at room temperature will not change the chirality at C-4, the absolute configuration of **1** is established as (2*S*,4*S*)-2-amino-4-methyl-hex-5-enoic acid.



The unambiguous synthesis of sterically pure **2** also establishes the stereochemistry of the naturally occurring amino acid, homoisoleucine, which had been isolated by Fowden and Smith [7] from *Aesculus californica*. Fowden *et al.* [8] had already proposed the 2*S*,4*S* configuration for homoisoleucine on the basis of a comparison of its solubility, ORD and CD with those of isoleucine.

## EXPERIMENTAL

Mps are uncorr. Optical rotations were determined on an ETL-NPL automatic polarimeter.

**Hydrogenation of 2-amino-4-methyl-hex-5-enoic acid (1).** 0.1 g **1** over Pt required one mole of hydrogen (15.4 ml) and gave (2*S*, 4*S*) 2-amino-4-methylhexanoic acid (**2**) (0.1 g) mp 240–246°; *m/e* 146, MH<sup>+</sup> from CI; [α]<sub>D</sub><sup>22</sup> +34.3° (c 0.385 in HOAc).

**(2*S*,4*S*)-2-Amino-4-methylhexanoic acid.** Refluxing 2*S*-(+)-1-bromo-2-methylbutane (4.5 g, prepared from 2*S*-(−)-2-methylbutan-1-ol) with ethyl acetamidomalonate (5.4 g) and NaOEt (0.6 g) in EtOH [3, 4] gave a mixture of (2*S*,4*S*) and (2*R*,4*S*) ethyl 2-acetamido-2-carboxy-4-methylhexanoate as a light yellow oil (5.6 g). Alkaline hydrolysis (10% NaOH, 2 hr), followed by refluxing (2 hr) of the acidified soln yielded a mixture of (2*S*,4*S*)

and (2*R*,4*S*)-2-acetamido-4-methylhexanoic acid (2.3 g), mp 190–191° from H<sub>2</sub>O, [α]<sub>D</sub><sup>27</sup> +5.1° (c. 2.86 in MeOH). The CI (isobutane) MS showed peaks at *m/e* 188, MH<sup>+</sup>, 142 (M<sup>+</sup>-COOH), 130 (M<sup>+</sup>-C<sub>4</sub>H<sub>9</sub>), 99 (M<sup>+</sup>-COOH-MeCO), 57 C<sub>4</sub>H<sub>9</sub><sup>+</sup>. An aq. soln of the above acid mixture was adjusted to pH 7 with ammonia and hydrolysed at 38° with powdered hog kidney acylase I (12 mg) [5, 6] overnight. The soln was acidified to pH 5, filtered, passed through a Zeocarb 225 cation exchange resin column in the H<sup>+</sup> form, and the column washed with H<sub>2</sub>O. The aq. eluates from the column were combined and evapd to dryness. Recrystallization of the solid from H<sub>2</sub>O gave (2*R*,4*S*) 2-acetamido-4-methylhexanoic acid (0.4 g), mp 192°. [α]<sub>D</sub><sup>30</sup> +22.3° (c. 6.27 in MeOH). Found: C, 58.0; H, 9.0; N, 7.3%; C<sub>6</sub>H<sub>17</sub>NO<sub>3</sub> requires: C, 57.7; H, 9.1; N, 7.5%. The amino acid was eluted from the column with 2*N* ammonia, and the soln concd to ca 200 ml. Cooling of the soln gave the lustrous, colourless crystals of (2*S*,4*S*)-2-amino-4-methyl-hexanoic acid (0.5 g), mp 240–244°. Found: C, 57.9; H, 10.4; N, 9.6%; C<sub>7</sub>H<sub>15</sub>NO<sub>2</sub> requires: C, 57.9; H, 10.4; N, 9.6%. [α]<sub>D</sub><sup>20</sup> +35.4° (c. 0.41 in glacial AcOH) and [α]<sub>D</sub><sup>22</sup> +25.7° (c. 1.0 in 5*N* HCl). The CI (isobutane) MS showed peaks at *m/e* 146 MH<sup>+</sup>, 100 (M<sup>+</sup>-COOH), 74 (M<sup>+</sup>-C<sub>4</sub>H<sub>9</sub>), and 57 C<sub>4</sub>H<sub>9</sub><sup>+</sup>, identical with the acid obtained on hydrogenation of **1** (MS, GLC, TLC, mp, mmp). This is in agreement with data recently obtained by Bernasconi *et al.* [9] who used α-chymotrypsin for the resolution of the C-2 centre. The 2*S* configuration of our amino acid was also confirmed by GLC of its *N*-TFA-*L*-prolyl methyl ester derivative [10].

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## REFERENCES

1. Rudzats, R., Gellert, E. and Halpern, B. (1972) *Biochem. Biophys. Res. Commun.* **47**, 290.
2. Gellert, E., Halpern, B. and Rudzats, R. (1973) *Phytochemistry* **12**, 689.
3. Albertson, N. F. and Tullar, B. F. (1945) *J. Am. Chem. Soc.* **67**, 502.
4. Snyder, H. R., Shekleton, J. F. and Lewis, C. D. (1945) *J. Am. Chem. Soc.* **67**, 310.
5. Fodor, P. J., Price, V. E. and Greenstein, J. P. (1949) *J. Biol. Chem.* **178**, 503.
6. Fodor, P. J., Price, V. E. and Greenstein, J. P. (1950) *J. Biol. Chem.* **182**, 467.
7. Fowden, L. and Smith, A. (1968) *Phytochemistry* **7**, 809.
8. Fowden, L., Scopes, P. M. and Thomas, R. N. (1971) *J. Chem. Soc. (C)*, 833.
9. Bernasconi, S., Corbella, A., Gariboldi, P. and Jommi, C. (1977) *Gazz. Chim. Ital.* **107**, 95.
10. Halpern, B. and Westley, J. W. (1965) *Biochem. Biophys. Res. Commun.* **19**, 361.

\* Part 3 in the series 'Constituents of a New Guinea *Boletus*'. For Part 2 see ref. [2].